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# Using quantitative magnetic resonance methods to understand better the gel-layer formation on polymer-matrix tablets

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Introduction: Magnetic resonance imaging is a powerful, non-invasive technique that can help improve our understanding of the hydrogel layer formed on swellable, polymer-matrix tablets, as well as the layer's properties and its influence on drug release.

Areas covered: In this paper, the authors review the NMR and MRI investigations of hydrophilic, swellable polymers published since 1994. The review covers NMR studies on the properties of water and drugs within hydrated polymers. In addition, MRI studies using techniques for determining the different moving-front positions within the swollen tablets, the polymer concentration profiles across them, the influence of the incorporated drug, and so on, are presented. Some complementary methods are also briefly presented and discussed.

Expert opinion: Using MRI, the formation of a hydrogel along with simultaneous determination of the drug's position within it can be observed non-invasively. However, the MRI parameters can influence the signal's intensity and therefore they need to be considered carefully in order to prevent any misinterpretation of the results. MRI makes possible an in situ investigation of swollen-matrix tablets and provides valuable information that can lead, when combined with other techniques, to a better understanding of polymeric systems and a more effective development of optimal dosage forms

Keywords: drug release, hydrogel, hydrophilic polymer, magnetic resonance imaging, matrix tablet, nuclear magnetic resonance, swelling

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#### 1. Introduction

Modified-release matrix tablets have been used extensively by the pharmaceutical industry as one of the most successful oral drug-delivery systems. The dominant matrix excipients for most modified-release formulations are hydrophilic polymers, which hydrate when in contact with the release medium. After the concentration of the medium is high enough, the glassy polymer transforms into a rubbery state and the polymeric molecules become flexible (i.e., the glass transition temperature  $T_{\rm g}$  of the polymer reduces to the system temperature T), forming a hydrogel layer that regulates the penetration of the medium into the matrix and the dissolution of the incorporated active ingredient. At the surface of the hydrogel layer some erosion may occur due to polymer dissolution through the chain disentanglements. As a result of all these processes running at the same time, different moving fronts have been postulated. The boundary between the hydrogel layer and the medium is called the erosion front. The diffusion front is an interface between the undissolved and the dissolved drug within the hydrogel layer. The boundary where

#### Article highlights.

- Drug release from hydrophilic matrix tablets is regulated by the hydrogel's formation around the dry matrix core. The hydrogel regulates the drug-release kinetics and the mechanism, therefore its thickness and properties should be known so as to be able to anticipate the drug-release dynamics.
- Nuclear magnetic resonance can provide useful information about the hydrogel's properties at different polymer concentrations in different conditions (medium's properties, at different temperatures, etc.).
- Magnetic resonance imaging is capable of providing, in a non-invasive way, internal images of the investigated materials and can therefore give information about moving-fronts' (penetration, swelling and erosion fronts) positions and the hydrogel's properties in situ. By combining the NMR parameters obtained from hydrogels at different polymer concentrations, the polymer concentration profiles across the formed hydrogel can be determined for different swelling times.
- The combination of MRI and USP-4 apparatus can simultaneously measure the polymer-swelling kinetics and the drug release, and can thus link together the hydrogel properties with the drug release.
- The combination of the data obtained with different methods can provide the information needed to design hydrophilic matrix tablets with the desired drug-release kinetics.

This box summarizes key points contained in the article

the glassy polymer transforms into the rubbery polymer is called the swelling front. Furthermore, there is an extra front that is frequently overlooked, referred to as the penetration front, which is the border between the dry glassy polymer and the hydrated glassy polymer, that is, the position of medium's penetration into the dry glassy polymer [1-3].

The hydrogel layer formed on polymer-matrix tablet is important for regulating the drug-release mechanism and kinetics from them. For this reason, numerous parameters have been investigated: the hydrogel layer's thickness and the position of the different fronts within the hydrogel [4,5], the presence of different excipients [6,7], the properties of the selected polymer and the investigated drug substance [8-10], the polymer concentration across the hydrogel, the mesh size of the polymeric network [11], and so on. A large spectrum of mathematical models describing the drug release from the hydrophilic matrix tablets has been developed, and these try to include many of these important parameters. However, in many cases the use of simple empirical or semiempirical models is sufficient; but, when reliable, detailed information is required, more complex, mechanistic theories must be applied [12].

Magnetic resonance imaging (MRI) is a non-invasive method that has the ability to provide internal images of soft materials such as hydrophilic matrix tablets. In addition, it can provide sufficiently high spatial (50 µm) and time resolution to monitor the swelling processes in situ. The purpose of this article is to review the current state-of-the-art for magnetic resonance measurements that are relevant to the swelling behavior of hydrophilic matrix tablets and to show the suitability of the technique for following the swelling kinetics and the release of the drug.

# 2. Brief summary of the techniques used for investigating the swelling process of polymer-matrix tablets

# 2.1 Studies using optical imaging

The swelling process of polymers has been studied using a variety of techniques. Optical imaging is perhaps the most well established of these. Studies of hydrogel-layer formation have been conducted using a variety of optical imaging methods, which cover techniques ranging from simple pictures of the matrix tablets' cross-sections to advanced video imaging with computer analyses.

Colombo and co-workers [4,13] and Bettini et al. [14] have identified the movement of the swelling, diffusion and erosion fronts in the established hydrating-gel layer using colored visual aids. In their method the tablet was placed between two transparent Plexiglas® sheets and the hydrogel formation was monitored from the axial direction of the sample. Simple video optical microscopy was introduced by Papadimitriou et al. to follow the swelling of hydroxypropyl methylcellulose (HPMC) in situ [15]. They found that the swelling is faster in the axial than in the radial direction as a consequence of the tableting process and relaxation. Gao and Meury introduced another optical-imaging method to determine the HPMC concentration profile across a swollen tablet in situ. On the basis of the empirical relationship between the scattered light intensity and the HPMC concentration for equilibrium swollen hydrogels, the polymer concentration profile of the gelled region was estimated. Using the postulated model they determined the position of the apparent hydrogel front, and in this way the hydrogel thickness [16]. Adler et al. [17] used fluorescence imaging, such as confocal laser scanning microscopy (CLSM), to monitor the swelling of internal domains within the hydrogel layer by tracking the embedded fluorescent particles, and a soluble fluorophore has been used to monitor liquid ingress into the HPMC matrices [18]. This technique offers good spatial resolution and sensitivity, with an optical sectioning capability and the time resolution necessary to monitor rapid changes. These are important in the early stages of swelling, where the rapid development of a coherent hydrogel diffusion barrier is a critical stage in the establishment of the extended release properties in hydrophilic matrices. The early stages of hydrogel-layer formation in situ were thus investigated by CLSM imaging using a cellulose-activated fluorophore, that is, Congo Red. Using this fluorophore, the mapping of the hydrated polymer regions during the development of the early hydrogel layer was possible. Their results revealed a temporal sequence in which the capillary uptake of the hydration medium into the tablet pores is followed



by a changing morphology of the surface of the HPMC particles, as they swell in a columnar manner to form the nascent gel barrier [19]. The same authors also investigated the increase in ionic strength on HPMC swelling and found that increasing the salt concentration in the hydration medium suppressed the hydrogel layer's growth. The method they proposed provides possible evidence in the form of images for mechanisms that may contribute to salt acceleration of drug release in HPMC matrices [19]. The CLSM technique was also used to investigate the influence of different sugars on early gel-layer formation and on accelerated drug release from HPMC matrices. It was shown that the presence of sugar has a different effect on HPMC particle hydration and swelling with regard to viscosity type. The authors found that the selection of diluents such as microcrystalline cellulose, fine particle size fraction of HPMC and its highest viscosity grade are key parameters for designing HPMC matrices with reduced sensitivity to high concentrations of dissolved sucrose [20-22]. Further, the effects of incorporated alkalizing buffers such as sodium citrate and tris(hydroxymethyl) aminomethane (THAM) on release of the weak acid drug from HPMC matrices were studied by CLSM imaging. The inclusion of each buffering system in HPMC tablets improved the release of drug in alkaline and acidic media. However, the inclusion of THAM was shown to maintain the pH elevation for a greater period and was found to have minimal effect on HPMC particle swelling and gel-layer formation [23,24].

There are also other potent imaging methods for investigating the mechanisms regulating drug release from the dosage forms. One of these is Fourier transform infrared (FTIR) spectroscopic imaging in attenuated total reflection (ATR). The ability to record spatially resolved chemical images as a function of time allowed the dynamic process to be viewed via simultaneous measurement of the distribution of the polymer, drug and water within the pharmaceutical formulations. However, a more detailed description of this method can be found in the cited literature and is out of the scope of this paper [25-27].

# 2.2 Studies using ultrasound

The formation of a hydrogel layer around the swellable tablets can be followed by a promising non-destructive method using ultrasound [28]. Luprano et al. used an ultrasound pulseecho technique to measure the hydrogel water sorption and monitored the advancement of the swollen-unswollen fronts of polymer films [29]. A further step was made by Leskinen et al., who monitored the erosion and swelling front movements during the tablet-dissolution process simultaneously, using an ultrasound window technique. It was noted that the sensitivity for following the hydrogel formation and thickening by ultrasound monitoring varied depending on the polymer under investigation. The polymers to be studied by ultrasound must possess certain acoustic properties in order to make the medium-polymer interface detectable. These properties also depend on the ultrasound frequency being

used. The authors found that multi-front detection is challenging because the hydrogels formed by different polymers may have totally different acoustic properties. Furthermore, owing to limited spatial resolution of these methods, observation of the early stages of polymer swelling can be problematic. In spite of these challenges, the ultrasound window technique introduced in their study has proved to be a promising method for simultaneous multi-front detection [30].

## 2.3 Studies using texture analyzer

An important characteristic of the formed hydrogel layer is its texture. This can be examined with texture analysis, a penetrometry technique where the material is subjected to a controlled force. From the resulting force-displacement plot, several parameters of the material can be derived that are directly related to the performance of the sample [31]. Although the texture analyzer technique is destructive and does not allow measurements in situ, the pharmaceutical applications for quality-control purposes have increased significantly during the past few years, as it is relatively simple, versatile and cost-effective; it is possible to use the same instrument for multiple measurements by changing either the testing probes or the measurement parameters [32]. Jamzad and co-workers studied the influence of water-soluble and insoluble excipients on the dynamics of hydration, front movement, erosion and drug release from HPMC matrix tablets containing a water-soluble drug. They determined the swelling front's position as well as the hydrogel layer's thicknesses. They found that within the context of hydrophilic polymeric matrices containing a water-soluble drug, excipients should not be regarded as neutral or simple additives because they are certainly capable of altering the water penetration, erosion, and hence the mechanism of drug release [6,33]. In addition to HPMC and PEO [6,33-34], xanthan-based tablets were also investigated using texture analysis [7,35]. It was found that the firmer matrix structure does not necessarily lead to a slower drug release because the hydrogel-layer thickness, polymer-network mesh size and the water distribution within the hydrogel are also important. It was concluded that only combinations of different techniques reveal the detailed structure of swollen matrix tablets that is necessary to understand the release of the active substance from them.

# 3. Short theory and principles of NMR and MRI

Nuclear magnetic resonance (NMR) is based on the phenomenon that nuclei possessing a magnetic moment (e.g., <sup>1</sup>H, <sup>19</sup>F, <sup>31</sup>P, <sup>13</sup>C, <sup>2</sup>H, etc.), after being placed in an external magnetic field, tend to align with the magnetic field, causing a weak net magnetization along the applied magnetic field. The application of a radiofrequency (rf) magnetic field alters the spin population distribution and consequently induces a small voltage in the surrounding rf coil by a process of electromagnetic induction



that generates the NMR signal. Unfortunately, the NMR signal is intrinsically weak, but it increases with increasing gyromagnetic ratio  $\gamma$  (i.e., a property that varies for different nuclei, and is largest for the hydrogen nucleus) and increasing magnetic field strength. As the rf pulse is turned off, the spins relax back to the equilibrium position owing to spin-lattice  $(T_1)$  and spinspin  $(T_2)$  relaxation processes (Figure 1). The NMR signal intensity thus depends on the concentration of the observed nuclei in the sample, on their relaxation times  $T_1$  and  $T_2$ , and on their self-diffusion coefficient (D) [36,37]. Using special NMR pulse sequences, the relaxation times  $T_1$  and  $T_2$  [38,39] and the selfdiffusion coefficient [40,41] of different materials can be measured. Those parameters can give valuable information about the local molecular environment and are thus implemented in research conducted on the different properties of matrix tablets, that is, information about the mobility of the water and polymer molecules and about water-polymer interactions in the swollen hydrogel can be obtained from the  $T_1$ ,  $T_2$  and Dmeasurements. With the application of more complex NMR pulse sequences, a better insight into water-polymer interactions can be provided [42-44].

Magnetic resonance imaging utilizes magnetic field gradients that are applied across the static magnetic field in order to obtain spatial information about the system. In the case of MRI, only the NMR signal from a small part of the sample (one pixel) is collected. To reach an adequate signalto-noise ratio for each pixel in acceptable times, a relatively high NMR signal is needed. Therefore, in most applications high magnetic fields are used and <sup>1</sup>H MRI is applied to samples containing <sup>1</sup>H nuclei in high concentrations and used to measure the distribution of the water within the investigated samples.

To observe the signal of all the protons in the sample, the TE (the time between the signal's excitation and detection) should be much shorter than their  $T_2$  and the repetition time (TR) should be long compared with their  $T_1$ . As the swelling of the polymer-matrix tablets is a dynamic process and the measuring time is limited to a couple of minutes, usually one-dimensional (1D) or two-dimensional (2D) experiments are performed, depending on the system symmetry. To increase the contrast between the media, hydrogel and dry polymer in the sample,  $T_1$ - or  $T_2$ -weighted magnetic resonance images, by changing TE and TR, respectively, or diffusion-weighted magnetic resonance images, by using a pulsed field gradient spin-echo (PGSE) technique, are applied [38]. Moreover, the relaxation times and self-diffusion maps can be used to obtain the spatial distribution of  $T_1$ ,  $T_2$ and D [38,45]. To improve the time resolution, it is also possible to use faster imaging methods, such as rapid acquisition with relaxation enhancement (RARE) [46-49].

Owing to the limited strength of the magnetic field gradients and their rising time and the use of shape pulses in 2D experiments, TE is in the range of a couple of milliseconds. In this time the NMR signal of the solid polymer has already decayed to zero and, by using standard

spin-echo MRI pulse sequences, only water protons with a long enough  $T_2$  can be spatially detected. For polymer and water protons with short T2 values (water in a hydrated glassy polymer and water in a hydrogel with a high polymer concentration), a special magnetic resonance sequence, the single-point-imaging (SPI) pulse sequence, which enables the imaging of protons with short  $T_2$  values and their modifications for spatial encoding of the relaxation times, can be used [50,51]. As the SPI sequence uses phase encoding in all the imaging directions it is time consuming, and, owing to the limited available measuring time during swelling, usually only a 1D SPI sequence can be applied. By combining different magnetic resonance techniques, the quantification of spatial properties during swelling, that is, the penetration, swelling and erosion front positions, and thus the hydrogel thickness together with the polymer concentration across the formed hydrogel can be determined.

In addition to the proton NMR signal, other nuclei can also be detected. If the drug substance incorporated into the matrix tablet contains one of those nuclei, the spatial distribution of the drug and thus the drug release during the polymer swelling can be concomitantly measured using MRI.

# 4. NMR spectroscopy as an analytical tool for the investigation of swollen polymer-matrix tablets

NMR can give valuable information about polymer properties in the rubbery state that can help in the search for an optimal polymer that will fulfil the specific needs of drug delivery from hydrophilic matrix tablets. The hydrogel's properties and their dependence on temperature changes, polymer concentrations, pH of the medium, drug incorporation, and so on, were studied using relaxation times and the self-diffusion coefficient.

The relaxation times  $T_1$  and  $T_2$  of the water protons are determined by the modulation of the intra- and intermolecular dipolar interactions, the magnetization transfer and the chemical exchange processes between the water and the polymer molecules. Therefore, the measurements of  $T_1$  and  $T_2$ allow a study of the physical and chemical properties of hydrogels [52]. The diffusion coefficient of the incorporated drug can give valuable information about the drug release, while the diffusion of the water is a measure of the waterpenetration rate, which determines the kinetics of the hydrogel's formation. In the hydrogels, both drug and water diffusivity can be measured, as the water and drug protons have slightly different NMR signals owing to a chemical shift. This makes it possible to distinguish the spectral lines of the water and the drug.

#### 4.1 Polymer and drug self-diffusion coefficient

It was found that the drug  $(D_D)$  and water diffusivity  $(D_{\rm W})$  decrease exponentially with increasing polymer concentration in the hydrogel, indicating an entangled



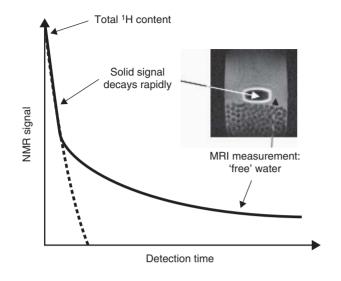


Figure 1. Schematic of basic NMR signal, the free induction decay after a radiofrequency pulse together with an example of a magnetic resonance image of a matrix tablet during swelling.

Adapted with permission from [74]

three-dimensional network structure [53-57] - except for egg albumin, where the relationship is linear and a diluted network structure was proposed [57]. The degree of  $D_{\rm D}$ decrease with polymer concentration depends on the degree of polymerization of the polymer, and is smallest for glucose (a monomer unit of HPMC) and largest for HMPC. Water diffusion, on the other hand, is independent of polymer chain length [53]. D<sub>D</sub> is also exponentially dependent on the drug concentration [53,56]. It was also observed that  $D_D$  and  $D_W$  do not vary significantly with molecular mass of the polymer [53-56]. The temperature dependence of  $D_D$  and  $D_W$  showed an Arrhenius-type behavior, with similar activation energies for the drug and water, comparable to the hydrogen-bonding energy. Based on these results, the authors concluded that the diffusion barrier for the drug and water is determined solely by the interactions between the water molecules [53]. It was also shown that the smaller drug is more mobile owing to the smaller size and fewer interactions with the polymer [58]. Study of  $D_D$  dependence on the drug's molecular mass showed that  $D_D$  strongly decreases with increasing molecular mass of the drug [59]. The results for the multicomponent gels (a mixture of HPMC, lactose and drug) showed that  $D_{\rm D}$  is affected by all the existing components, resulting in an additive retarding effect from all the components [53].

From the NMR measurements of  $D_D$  and  $D_W$  in the presence of various polymers and drugs, it was concluded that medium diffusion depends on the interaction between the medium and the matrix polymer [54], and that the  $D_{\rm D}$  is dictated by the microviscosity of the system and by the steric obstruction mechanism [53,55]. Based on a comparison of the measured data with the theoretical approach, agreement with the free-volume theory [60,61], predicting that drug diffusion is the consequence of jumps in voids, which are created by correlated motions between the water and the surrounding polymer molecules, was confirmed [53,55]. Some deviation from the free-volume theory was observed for the diffusion of larger molecules at higher polymer volume fractions, where other models were proposed instead [54]. A comparison between experimental values with theoretical models gives information about the hydrogel's microstructure connected with the drug's mobility in the hydrogel and thus the drug-release mechanism.

#### 4.2 NMR relaxation times

The temperature dependences of the proton-relaxation times  $T_1$  and  $T_2$  in the hydrogels were used to determine the transition temperature from the glassy to the rubbery state,  $T_{\rm g}$ . The spin-spin relaxation time  $T_2$  is almost constant below  $T_{\rm g}$  and increases significantly with temperature above  $T_{\rm g}$ . The spin-lattice relaxation time  $T_1$  has a minimum at  $T_2$ . The  $T_{\rm g}$  values that were determined were verified by the values obtained from a differential scanning calorimetry (DSC) experiment [62-64].

The relaxation times were also used to investigate the water mobility within the hydrogels, to obtain information about the hydrogel's structure (the homogeneity of the hydrogel) and to measure the average number of water molecules bound per polymer repeating unit. In the hydrogel's measured relaxation times there are two exponentials, indicating two types of proton: a 'polymer' pool, comprising non-exchangeable polymer protons ( $T_1 \sim 250$  ms and  $T_2$  in the order of hundreds of microseconds); and a 'water' pool, comprising protons of bound and free water and exchangeable polymer protons. The latter are in a fast exchange relative to the NMR timescale, having a single component of  $T_1$  and  $T_2$  ( $T_1$  from hundreds of milliseconds to seconds and  $T_2$  in the range from some to hundreds of milliseconds). The relaxation times of the water pool decrease with increasing polymer concentration [49,65-71], reflecting the increase of interactions between the water and the polymer molecules with increasing polymer concentration. The relaxation times of the polymer pool are, on the other hand, independent of the polymer concentration in the hydrogels of poly(amidoamine)s, suggesting that the mobility of the polymer is relatively constant once the polymer is in contact with the water [65]. By contrast, in the HPMC hydrogels, significant changes in the mobility of the polymer were observed when changing the polymer concentration [69,72].  $T_1$  and  $T_2$  were found to be independent of the polymer's molecular mass, but  $T_1$  was sensitive to the polymer substitution type in different cellulose ethers [66,67]. From  $T_1$  and  $T_2$  data it is also possible to calculate the average number of water molecules bound per polymer repeating unit. It was found that this is independent of the molecular mass within a given polymer type, but increases with the

degree of hydrophilic substitution of the polymer chains [66]. The  $T_1$  and  $T_2$  parameters are influenced by polymer crosslinking as well. Calucci et al. observed a decrease of both relaxation times with decreasing degree of crosslinking of the hydrogel, indicating that higher crosslinking leads to a more rigid heterogeneous polymer structure [65]. A study of the impact of the medium's pH and ionic strength on the hydrogels made of xanthan, which is an anionic polymer and thus its properties are strongly affected by pH and ionic strength, on the relaxation times and on the mobility of the protons within the hydrogel, showed that  $T_1$  is mostly independent with respect to different media, whereas  $T_2$  relaxation times are significantly lower using pH 1.2 media than for media with higher pH values. This was interpreted as a more restricted mobility of the polymer chains in low-pH media, as the spin-spin relaxation time is strongly correlated with the rigidity of the hydrogel – a slower  $T_2$  decrease, showing a more rigid hydrogel, which was confirmed by the rheological studies [68]. The results obtained by NMR for different hydrogels are summarized in Table 1.

# 5. What can be seen within swollen-matrix tablets using MRI?

To obtain spatial information about the tablet's swelling and erosion, magnetic resonance imaging has been used extensively [73-75]. The technique is non-invasive, fast enough to be able to follow changes during the swelling processes and can give reliable information about the medium's penetration into the matrix tablet and hydrogel formation.

#### 5.1 MRI methods and equipment

Different approaches have been applied to determine the erosion, swelling and penetration fronts and thus the hydrogel thickness during swelling of the matrix tablet. One of the most frequently used approaches is that of  $T_1$ -weighted images. In the hydrogel-forming systems the magnetic resonance signal of the hydrogel is higher than the signal of the medium on the  $T_1$ -weighted images owing to the longer  $T_1$ of the medium. Furthermore, the MRI signal increases when the polymer transforms from the glassy (the  $T_2$  of which is too short to be observed by a spin-echo MRI sequence) to the rubbery state owing to the longer  $T_2$  of the hydrogel [67,68,76-86]. Therefore, the position of the swelling front is usually determined by the appearance of the MRI signal inside the tablet and the position of the erosion front is determined at the position where the signal decreases to the value of the magnetic resonance signal of the bulk medium (Figure 2A). The difference between the swelling front and erosion front position represents the thickness of the hydrogel, which is an important parameter influencing drug release from the hydrophilic matrix tablets.

Another approach to determining the hydrogel thickness is to measure the spatial variation of the spin-spin relaxation time, that is, the  $T_2$  map [49,58,68-70,79,87-94]. Combining the

MRI technique with a Carr-Purcell-Meiboom-Gill (CPMG) sequence does not significantly increase the measurement time compared with the standard spin-echo technique.  $T_2$ was found to be changed through the hydrogel, from very short values at the interface between the glassy and rubbery regions to much longer values at the interface between the rubbery hydrogel and the bulk region (Figure 2B). As  $T_2$ reflects the rotational and translational freedom of the water molecules, the changes in  $T_2$  over the hydrogel regions indicate that the hydrated polymer in the hydrogel layer close to the glassy state possesses a greater proportion of more tightly bound water and that a strong interaction occurs between the adsorbed water and the polymer [88,91-93]. By using spinecho-based MRI pulse sequences only protons with long enough  $T_2$  values compared with TE can be observed. To overcome the problem of the short  $T_2$  values of the hydrogels with high polymer concentrations, the SPI technique that enables samples with short  $T_2$  values to be imaged was used [68,72]. As the SPI technique is time-consuming, only 1D imaging can be performed in order to obtain a sufficiently high time resolution. It was shown that with a combination of the  $T_1$ -weighted spin-echo MRI technique, the SPI technique and  $T_2$  mapping measured with the SPI technique, an accurate determination of the medium-penetration position, the position of hydrogel formation, that is, the swelling front and the erosion front, can be achieved for polymers, even though the  $T_2$  values for hydrogels at high polymer concentrations are short (Figure 3) [68]. To study directly the polymer response during the medium penetration and swelling, D<sub>2</sub>O was used as the penetration medium. With this approach only the polymer protons contribute to the <sup>1</sup>H signal, and thus direct measurements of the molecular processes of the polymer carriers during swelling can be imaged [72,95].

MRI measurements of water self-diffusion coefficient maps during swelling are also a very useful technique for obtaining the properties of the hydrogel with different polymer concentrations formed during swelling [49,76,77,81,87-89,91-93,96-97]. For different polymers it was shown that the water diffusivity is restricted in the polymer matrix owing to the limited mobility of the polymer chains, and that the diffusion coefficient changes with distance from the dry polymer through the hydrogel towards the medium region as the polymer concentration changes through the hydrogel layer. The water selfdiffusion coefficient was found to have a constant value near the polymer-water interface and was similar to the diffusion coefficient of free water, indicating that in this region a dilute solution of polymer chains exists for all the swelling times. Water self-diffusion is much slower inside the tablet during the early stages of swelling, where the diffusion coefficient changes with the swelling time as the water penetrates the tablet and causes an increase in mobility of the polymer chains [76,77,88].

The intensity of the MRI signal increases with magnetic field strength, and therefore in most cases high fields were used in order to achieve better resolution and a higher



Table 1. Summary of the NMR results obtained from measuring the spin-lattice  $T_1$ , spin-spin  $T_2$  relaxation times, water and drug self-diffusion coefficients of the hydrogels made from different polymers and drugs forming the matrix tablets.

NMR parameter	Polymer	Drug	Observations and conclusions	Ref.
$D_{W}$	HPMC PVA, HPMC, PNNDEA, PNIPA		Polymer concentration: exponential dependence	[53,56-57] [54]
	EA HPMC HEC, HPC		Polymer concentration: linear dependence Polymer weight: no significant changes	[57] [53,54,56] [55]
	HPMC, glucose, lactose		Polymer chain length: not dependent	[53]
	HPMC		Temperature: Arrhenius-type behavior	[53]
$D_{D}$	HPMC	Adinazolam	Polymer concentration: exponential	[53]
	HPMC, HEC, HPC	Sodium salicylate	dependence	[55]
	HPMC	Naproxen sodium	'	[56]
	HPMC	Adinazolam	Drug concentration: exponential dependence	[53]
	HPMC	Naproxen sodium		[56]
	HPMC	Adinazolam	Polymer molecular mass: no significant changes	[53]
	HPMC	Naproxen sodium		[56]
	HPMC, HEC, HPC	Sodium salicylate		[55]
	HPMC	Adinazolam	Polymer chain length: longer chains $\Rightarrow$ smaller $D_D$	[53]
	HPMC	Adinazolam	Temperature: Arrhenius-type behavior	[53]
	PEG	FITC-dextrans	Drug molecular mass: higher-molecular- mass ⇒ more restricted mobility	[59]
	HPMC	5-Fluorouracil or triflupromazine-HCl	Drug size: smaller drug is more mobile	[58]
$T_1$ and $T_2$	PAAHn		Polymer concentration: higher polymer	[65]
	HPMC, HEC, HPC HPMC XAN XAN + locust bean gum		concentration $\Rightarrow$ shorter $T_1$ and $T_2$	[66,67] [49,58,69-70 [68] [71]
	HPMC		Polymer molecular mass: no influence	[66,67]
	HPMC, HEC, HPC		Polymer substitution: $T_1$ sensitive, $T_2$ no influence	[66,67]
	PAAHn		Polymer crosslinking degree: lower crosslinking degree $\Rightarrow$ shorter $T_1$ and $T_2$	[65]
	XAN + locust bean gum	Mannitol as model drug	Incorporated drug and its solubility: $T_1$ no influence, $T_2$ slightly increases	[71]
	XAN	model drug	Medium pH: $T_1$ not dependent, lower pH $\Rightarrow$ shorter $T_2$	[68]

DCP: Dicalcium phosphate; EA: Egg albumin; FITC-dextrans: Fluoresceine isothiocynate-labeled dextrans; HEC: Hydrohyethylcellulose; HPC: Hydroxypropylcellulose; HPMC: Hydroxypropyl methylcellulose; MCC: Microcrystalline cellulose; PAAH: Poly(amidoamine); PEG: Poly(ethylene glycol); PEO: Polyethylene oxide; PNIPA: Poly (N-isopropylacrylamide); PNNDEA: Poly(N,N-diethylacrylamide); PVA: Poly(vinyl alcohol); PVP: Poly(vinylpyrrolidone); XAN: Xanthan.

sensitivity. However, as high-field MRI systems are very expensive, their use is limited. To expand the use of MRI in pharmaceutical research, a low-field, bench-top MRI system that is accessible for standard laboratory environment was developed. It proved to have sufficient resolution to monitor tablet hydration and swelling [98-101].

As one of the most important properties of tablets with controlled drug release is time dependence of the drug delivery itself, instantaneous measurement of the formed hydrogel's properties and drug release would provide very useful information. For that reason, a combination of MRI and the USP-4 apparatus was developed (Figure 4) [78,79,86,102].

The other approach to concomitantly measuring the released drug was to measure the <sup>1</sup>H spectrum of the medium. Owing to the chemical shift, the drug's signal can be distinguished from the medium's signal [72]. The third option is to measure the drug's signal directly; for example, by measuring the <sup>19</sup>F MRI signal [58].

#### 5.2 Results and findings obtained by MRI

The dependences of the relaxation times and the diffusion coefficient on the polymer's concentration in the hydrogel were used to determine the polymer's concentration profile in the hydrogel during swelling, which directly shows the

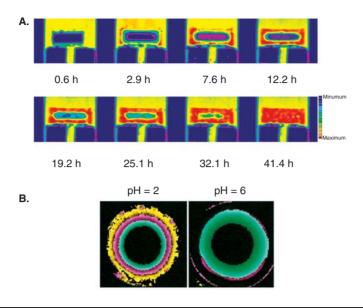


Figure 2. Representative examples of the <sup>1</sup>H magnetic resonance images. A. T<sub>1</sub>-weighted magnetic resonance images of a Contramid-I tablet at different swelling times with an in-plane resolution of 78 µm and a slice thickness of 0.5 mm. B. Effective  $T_2$  maps of HPMC tablets at 190 min after immersion in a medium of pH 2 and a medium of pH 6 with an in-plane resolution of 117  $\mu$ m and a slice thickness of 2 mm. The effective  $T_2$ s range from 440 ms (yellow color) to 0 ms (black). Adapted with permission from [79] A. [84].

rate of water penetration into the polymer matrix and thus the swelling kinetics. By knowing the dependence of the polymer hydrogel's concentration on the relaxation times and by using a phantom sample with known relaxation times to calibrate the signal, the polymer concentration across the formed hydrogel can be determined from the signal intensity for each pixel in the image [49,67]. Another method to determine the polymer-concentration profiles as functions of the distance and the swelling time is to measure  $T_1$  [95],  $T_2$  [49,58,69,70,89] or the diffusion maps [85,103]. However, the polymer concentration within the hydrogel cannot be determined in a straightforward way from the MRI data and special care should be taken in the evaluation of the data. Namely, the presence of trapped air in the tablet can lead to an overestimation of the polymer concentration as a result of the volume occupied by the air and because of shortening of water  $T_2$  in the vicinity of the air resulting from the susceptibility effects [89]. Further, using the spinecho method it is not possible to determine small water concentrations in the hydrogel because of the effect of  $T_2$  and diffusion losses [70].

From the different MRI methods, various findings for polymer-matrix tablets were observed that can lead to a better understanding of the swelling process and drug release. Based on these fundamental results, optimum polymer selection for the required performance of the drug-delivery system is possible. One interesting study involved NMR measurements of water ingress into polymer powders with different levels of compaction and, therefore, with different levels of porosity. On the basis of the NMR results the authors proposed a

mechanism of water penetration into the solid polymer matrix. It was shown that at the interface between the water and the polymer, first the vapor diffuses into the polymer, followed by liquid water diffusion, and that the liquid-water transport rate is controlled by the watervapor diffusion between the powder particles [96,104]. This was supported by other measurements where the core expansion of HPMC matrix tablets was observed before hydrogel formation [49,70,88,89,94]. MRI showed that the tablets' properties during swelling are different in the axial direction from in the radial direction [49,81,84,85,88,89,91,94,100]. This was explained by release of the compression stress from the granules or powder particles as the water penetrates the matrix, or with a smaller amount of bound water in the radial direction leading to different hydrogel properties [88,91,94].

It is not only the thickness, but also the hydrogel's properties that have an important role in the control of drug release. Using MRI, it was shown that the polymer's characteristics (different substitution type, molecular mass, hydrophilicity, polymer-water and polymer-polymer interactions) influence the hydrogel's thickness, on the one hand, and the amount of absorbed medium, on the other, leading to different hydrogel properties and a different drug release rate [56,57,67,76-78,92-93]. Furthermore, the influence of different polymer crosslinking of high amylose starch on the swelling was investigated, showing that higher crosslinking leads to higher swelling [82].

Usually, polymer-matrix tablets are investigated by MRI without any incorporated drug because of the complexity of the experiments. However, some valuable results were



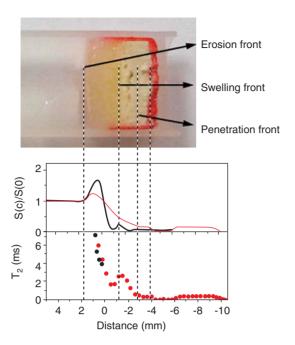


Figure 3. Photographs of a hydrated xanthan tablet after 3 h of swelling in a pH 1.2 medium, together with 2D spinecho MRI (black line) and 1D SPI (red line) normalized signal intensities and  $T_2$  profiles ( $T_2$  values were determined from 2D  $T_2$  maps for  $T_2$  longer than 5 ms (black circles) and from 1D SPI measurements for shorter  $T_2$  values (red circles)). The penetration front was determined from the increase of the normalized single-point-imaging signal intensity above the signal intensity of the dry tablet, the swelling front from the  $T_2$  value of the maximally hydrated glassy polymer (2.6 ms in the xanthan tablet) and the erosion front from the decrease of the 2D spin-echo MRI signal.

Adapted with permission from [68]

obtained from studying the influence of an incorporated drug on the swelling behavior of polymer-matrix tablets. It was shown that the presence of the drug increases the hydrogel's thickness at drug concentrations that are high enough and that highly soluble drugs have a greater impact than a drug with a lower solubility [93,94]. The presence of the drug also increases the medium penetration rate; however, this is more in the case of a more highly soluble drug than a lower solubility drug [81,94]. The rate of erosion was also found to depend on the drug's solubility. It is much slower for a lowsolubility drug [94]. As mentioned already, direct measurement of the drug's position in the hydrogel during swelling is possible when the drug molecule possesses other MRI-seen nuclei, such as <sup>19</sup>F. It was shown that the drug self-diffusion coefficient should be higher than the expansion rate of the matrix tablet in order to enable the release of the drug from the polymer tablet [58]. MRI was also used to examine the floating properties of different matrix tablets in the presence of gas-generating substances or substances with a low density [99,100].

The influence of external parameters, such as the temperature, the medium's pH, the presence of air bubbles, stirring of the medium and the tablet size, on the swelling behavior was also studied extensively using MRI. It was found that a higher temperature increases the water-penetration rate and can change the water diffusion from case II to Fickian behavior [79,82,105]. The temperature dependence of the swelling rate can be described by the Arrhenius law with an activation energy similar to the potential energy of the H-bonds, indicating that the swelling is a consequence of the breaking up of the H-bonds [83]. The medium's pH also causes different swelling or water-penetration mechanisms with respect to the polymer matrix. For HPMC polymers, it was found that the diffusion changes from Fickian at pH 6 to case II in the presence of a medium with a pH equal to 2 [79]. In xanthan the medium's pH does not influence the penetration and swelling front positions, but it lowers the erosion rate at low pH and therefore influences the resulting thickness of the hydrogel [68]. The tablet size was shown to influence the swelling behavior, that is, in the case of smaller tablets an increased swelling rate and water uptake were observed [84]. It was shown that a higher stirring rate causes a more rapid medium penetration and faster polymer erosion [85]. Furthermore, the flow of the medium through the cell containing the tablet was studied, leading to a thinner hydrogel due to mechanical perturbation of the hydrogel that strips away the outermost, highly hydrated, polymer chains [86]. The observed influences of the polymer type, incorporated active ingredients and external parameters on the swelling kinetics are summarized in Table 2. From a comparison of the hydrogel thickness and drug release, the MRI measurements show that the hydrogel thickness regulates the drug release, that is, drug release is faster in the case of a thinner hydrogel layer [68,78,88]. However, as the hydrogel-layer thickness is a consequence of different polymer conformations, the hydrogel's microstructure is also equally important. MRI measurements were also used to verify mathematical models describing the swelling and drug release from polymer-matrix tablets and to determine the critical parameters that can sometimes be the key to a successful design of matrix tablets for a particular application [76,77,96,103,106-107].

# 6. Expert opinion

Drug release from hydrophilic polymer-matrix tablets is a very complex process that is mainly influenced by the structure of the hydrogel layer [1]. Magnetic resonance is a very useful technique for the study of the hydrogel's properties and its thickness because it is non-invasive, sufficiently fast to be able to follow the changes during the swelling processes, and can give reliable information about the medium's penetration into the matrix tablet and the hydrogel's formation. In addition, it has the advantage of not requiring any contrast agents to obtain the system properties. Other methods are also used in the pharmaceutical research of the swelling of polymer



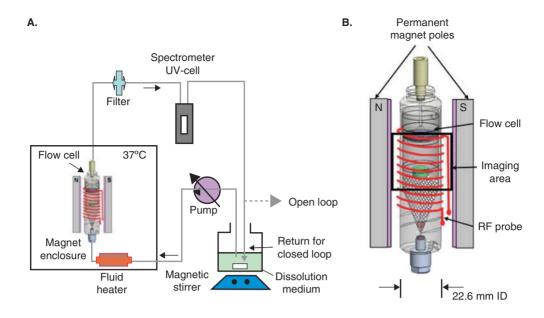


Figure 4. Schematic diagram of (A) a low-field magnetic resonance imager with an integrated USP-4 dissolution circuit (MARAN-iP) and (B) an enlarged picture of the MRI-compatible flow cell. Adapted with permission from [74].

Table 2. Summary of the results obtained from in situ MRI measurements of the matrix tablets made from different polymers and drugs swelling in media with various pH values and temperatures.

Processes and parameters during polymer swelling	Polymer type/drug or additive	Observations and conclusions	Ref.
Medium penetration	HPMC, PVA PEO	Depends on the polymer molecular masses: higher molecular mass ⇒ slower medium penetration rate	[76,91] [85]
	High-amylose starch	Tablet size: smaller size ⇒ faster penetration and larger water uptake	[84]
	High-amylose starch PMMA	Temperature: Arrhenius-type behavior	[82,83] [105]
	HPMC XAN	Medium pH: lower pH ⇒ faster penetration No influence Drug increases the penetration rate:	[79] [68]
	High-amylose starch/ciprofloxacin or acetaminophen	higher drug molecular mass $\Rightarrow$ faster penetration	[81]
	HPMC/mannitol or DCP PEO	higher soluble drug ⇒ faster penetration Stirring rate: higher stirring rate ⇒ faster penetration	[94] [85]
Erosion	XAN HPMC/mannitol or DCP PEO	Medium pH: lower pH ⇒ slower erosion  Drug: higher soluble drug ⇒ faster erosion  Stirring rate: higher stirring rate ⇒ faster erosion	[68] [94] [85]
Gel thickness	HPMC, HEC, HPC High-amylose starch PEO HPMC	Polymer substitution: higher hydrophilicity ⇒ thicker gel Crosslinking: higher crosslinking ⇒ higher swelling Stirring rate: higher stirring rate ⇒ thinner hydrogel Flow-through: causes thinner hydrogel	[67] [82] [85] [86]

DCP: Dicalcium phosphate; HEC: Hydrohyethylcellulose; HPC: Hydroxypropylcellulose; HPMC: Hydroxypropyl methylcellulose; PEO: Polyethylene oxide; PVA: Poly (vinyl alcohol); XAN: Xanthan.



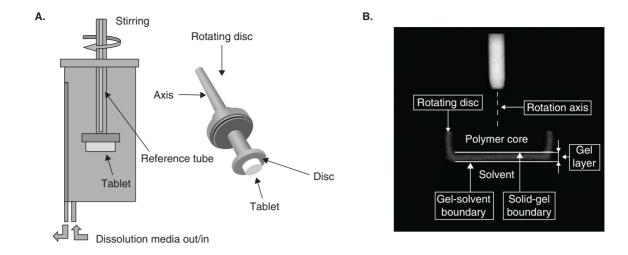


Figure 5. Combination of a rotating disc dissolution and MRI experiment. A. Schematic drawing of the MRI stirring release cell. B. The magnetic image obtained from a rotating disc dissolution experiment, showing the gel-solvent, the solid-gel boundary and gel layer.

Adapted with permission from [72] A. [85].

tablets, but these methods can be destructive and do not allow the in situ measurements (texture analyzer), need the addition of different markers that can influence the properties of the hydrogel layer (optical imaging), or cannot accurately distinguish between different moving-front positions (ultrasound, X-ray microtomography). Nevertheless, some of these are more accessible, cheaper and easier to use, and it is important to stress that they provide complementary information about the properties of the matrix tablets.

The applicability of MRI in pharmaceutical research has been known for a long time [73], but the optimal procedure for time and spatial resolution for observing processes occurring during swelling and drug release is still under investigation. One of the main obstacles in MRI studies is that a quantitative characterization of the MRI results of a particular polymer system is not straightforward, because the NMR parameters for hydrogels with different polymer concentrations should be determined first - a very labor-intensive part of the research. Another problem is also the short  $T_2$  of the polymer and hydrogel with a high polymer concentration, which cannot be observed using the standard spin-echo MRI technique. This can lead to imprecise determination of the swelling front and consequently an underestimation of the hydrogel layer's thickness - a weaknesses that many researchers are unaware of. A possible solution was shown to be the use of the SPI sequence [68,72], but it is time-consuming and, owing to unacceptable time resolution, only 1D images can be determined, which requires the use of a special sample geometry. The other possibility would be to use the Sweep Imaging with Fourier transform (SWIFT) technique, which is sensitive to nuclei with short  $T_2$  values and is significantly faster than the SPI technique [108]. With these methods it is possible to determine precisely the penetration front, but the problem remains: how

does one detect the position of the hydrogel formation, meaning the true swelling front's position, where the polymer transforms from the glassy to the rubbery state? However, it was proposed that a combination of the SPI signal intensity and the  $T_2$  measurements could overcome this problem [68]. Another obstacle to the MRI measurements is the diffusion losses causing a loss of the MRI signal and, therefore, an underestimation of the  $T_2$  values, which leads to an overestimation of the polymer concentration across the hydrogel layer. These effects should be considered when the polymer concentration profiles during tablet swelling are determined. The magnetic resonance images are strongly affected by selected imaging parameters and pulse sequences, and because the hydrated polymeric systems are very complex, interpretation of the magnetic resonance images can sometimes be misleading if the magnetic resonance parameters of the investigated systems are not considered properly.

A further step in the MRI implementation of research on the polymer-matrix tablets is simultaneous measurements of the behavior of the hydrogel layer's formation and the drug's position within it. This can be achieved by observing other nuclei (such as <sup>19</sup>F) that are part of the drug molecule, but are not present in the composition of the polymer or the medium [58]. The drawback is that the NMR sensitivity of such a nucleus is much lower than for the protons, which require a high magnetic field and a high ratio of the observed nuclei in the drug molecule. Drug release can also be monitored simultaneously with the MRI measurements using flow-through systems, where only the released drug can be detected using standard UV-VIS spectroscopy [74]. Although this method does not allow any observation of the drug's position within the hydrogel, it can give useful information about the hydrogel's impact on the drug release.

One of the reasons for the scarce use of MRI in pharmaceutical research is the high investment and running costs of superconducting MRI systems. Therefore, low-field MRI, bench-top instruments were developed recently. Their lower price and running costs make their widespread use possible in analytical laboratories. The use of these systems is partially limited by the lack of sensitivity for detailed investigations of the hydrogel's properties and the drug's position. However, on the other hand a bench-top MRI system is still very useful in the optimization process for matrix-tablet formulation and is expected to be developed further.

Most of the MRI research on swelling tablets was performed using in vitro methods, with the intention to predict the behavior of the systems in vivo. However, a major concern with in vitro experiments is the lack of a direct correlation between the in vitro and in vivo conditions. In some cases attempts to simulate the in vivo conditions were made by changing the medium's pH and ionic strength [68,79], fixing the temperature during swelling to 37°C [78,81-84,87,94,102], implementing the USP-4 flowthrough cell [78,86,102], or introducing a specially designed stirring cell (Figure 5) [85] to simulate the mechanical stress in the gastrointestinal tract. However, the simulated conditions are never exactly the same as the physiological ones, and some improvements should be made in the direction of simulating the influence of food, gender, age and disease conditions [109]. In vivo monitoring of the tablet's position

can be achieved by the incorporation of MRI contrast agents into the delivery system. Some attempts have already been made in vivo to follow the position of a floating tablet in the stomach [110].

It can be concluded that the MRI technique is a complementary method for other methods commonly used in pharmaceutical research. It can provide extra information that can lead, together with the results of other techniques, to a better understanding of polymeric systems and, therefore, to the design of systems with the desired properties for a particular need. The technique is still developing and some further improvements are expected to make MRI even more accurate and also more accessible for a wider range of applications in pharmaceutical research and analytics.

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#### **Declaration of interest**

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#### **Bibliography**

Papers of special note have been highlighted as either of interest (•) or of considerable interest ( • • ) to readers.

- Colombo P, Santi P, Bettini R, et al. Drug release from swelling-controlled systems. In: Wise DL, editor, Handbook of pharmaceutical controlled release technology. Marcel Dekker, Inc., New York; 2000
- Brazel CS, Peppas NA. Modeling of drug release from swellable polymers. Eur J Pharm Biopharm 2000;49:47-58
- Ferrero RC, Bruneau N, Barra J, et al. 3 Hydrophilic cellulose derivatives as drug delivery carriers: the influence of substitution type on the properties of compressed matrix tablets. In: Wise DL, editor, Handbook of pharmaceutical controlled release technology. Marcel Dekker, Inc., New York; 2000
- Colombo P, Bettini R, Peppas NA. Observation of swelling process and diffusion front position during swelling in hydroxypropyl methyl cellulose (HPMC) matrices containing a soluble drug. J Control Release 1999;61:83-91
- 5. Kazarian SG, van der Weerd J. Simultaneous FTIR spectroscopic imaging and visible photography to monitor tablet dissolution and drug release. Pharm Res 2008;25:853-60
- Pillay V, Fassihi R. A novel approach for constant rate delivery of highly soluble bioactives from a simple monolithic system. J Control Release 2000;67:67-78
- Baumgartner S, Pavli M, Kristl J. Effect 7. of calcium ions on the gelling and drug release characteristics of xanthan matrix tablets. Eur J Pharm Biopharm 2008;69:698-707
- Baumgartner S, Planinsek O, Srcic S, et al. Analysis of surface properties of cellulose ethers and drug release from their matrix tablets. Eur J Pharm Sci 2006;27:375-83
- Talukdar MM, Michoel A, Rombaut P, et al. Comparative study on xanthan gum and hydroxypropylmethyl cellulose as matrices for controlled-release drug delivery I. Compaction and in vitro drug release behaviour. Int J Pharm 1996;129:233-41
- Ferrero C, Massuelle D, Doelker E. Towards elucidation of the drug release mechanism from compressed hydrophilic matrices made of cellulose ethers. II.

- Evaluation of a possible swelling-controlled drug release mechanism using dimensionless analysis. J Control Release 2010;144:223-33
- 11. Baumgartner S, Kristl J, Peppas NA. Network structure of cellulose ethers used in pharmaceutical applications during swelling and at equilibrium. Pharm Res 2002;8:1084-90
- Siepmann J, Peppas NA. Modeling of drug release from delivery systems based on hydroxypropyl methylcellulose (HPMC). Adv Drug Deliv Rev 2001;48:39-157
- 13. Colombo P, Bettini R, Massimo G, et al. Drug diffusion front movement is important in drug-release control from swellable matrix tablets. J Pharm Sci 1995;84:991-7
- 14. Bettini R, Catellani PL, Santi P, et al. Translocation of drug particles in HPMC matrix gel layer: effect of drug solubility and influence on release rate. J Control Release 2001;70:383-91
- 15 Papadimitriou E, Buckton G, Efentakis M. Probing the mechanisms of swelling of hydroxypropylmethylcellulose matrices. Int J Pharm 1993;98:57-62
- 16. Gao P, Meury R. Swelling of hydroxypropyl methylcellulose matrix tablets. 1. Characterization of swelling using a novel optical imaging method. J Pharm Sci 1996;85:725-31
- Adler J, Jayan A, Melia CD. A method for quantifying differential expansion within hydrating hydrophilic matrices by tracking embedded fluorescent microspheres. J Pharm Sci 1999;88:371-7
- Melia CD, Marshall P, Stark P, et al. Investigating in vitro drug release mechanisms inside dosage forms - Monitoring liquid ingress in HPMC hydrophilic matrices using confocal microscopy. Adv Exp Med Biol 1997;423:129-35
- Baiwa GS, Hoebler K, Sammon C, et al. 19. Microstructural imaging of early gel layer formation in HPMC matrices. J Pharm Sci 2006;95:2145-57
- Williams HD, Ward R, Hardy IJ, et al. The extended release properties of HPMC matrices in the presence of dietary sugars. J Control Release 2009;138:251-9

- Williams HD, Ward R, Culy A, et al. 21. Designing HPMC matrices with improved resistance to dissolved sugar. Int J Pharm 2010;401:21-59
- Williams HD, Ward R, Hardy IJ, et al. The effect of sucrose and salts in combination on the drug release behaviour of an HPMC matrix. Eur J Pharm Biopharm 2010;76:133-436
- Pygall SR, Kujawinski S, Timmins P, et al. Mechanisms of drug release in citrate buffered HPMC matrices. Int J Pharm 2009;370:110-20
- 24. Pygall SR, Kujawinski S, Timmins P, et al. The suitability of tris (hydroxymethyl) aminomethane (THAM) as a buffering system for hydroxypropyl methylcellulose (HPMC) hydrophilic matrices containing a weak acid drug. Int J Pharm 2010;387:93-102
- 25. Kazarian SG, Kong KWT, Bajomo M, et al. Spectroscopic imaging applied to drug release. Food Bioprod Process 2005;83:127-35
- 26 Kazarian SG, Chan KLA. Applications of ATR-FTIR spectroscopic imaging to biomedical samples. Biomembranes 2006;1758:858-67
- Velasco D, Danoux CB, Redondo JA, et al. pH-sensitive polymer hydrogels derived from morpholine to prevent the crystallization of ibuprofen. J Control Release 2011;149:140-5
- Konrad R, Christ A, Zessin G, et al. The use of ultrasound and penetrometer to characterize the advancement of swelling and eroding fronts in HPMC matrices. Int J Pharm 1998;163:123-31
- Luprano VAM, Montagna G, Molinas B, et al. Glass-rubber phase transformation detected in polymers by means of ultrasonic waves. J Alloys Compd 2000:310:382-7
- Leskinen JTT, Hakulinen MA, Kuosmanen M, et al. Monitoring of swelling of hydrophilic polymer matrix tablets by ultrasound techniques. Int J Pharm 2011;404:142-7
- Durig T, Fassihi R. Guar-based 31. monolithic matrix systems: effect of ionizable and non-ionizable substances and excipients on gel dynamics and release kinetics. J Control Release 2002;80:45-56



#### Using quantitative magnetic resonance methods to understand better the gel-layer formation on polymer-matrix tablets

- Li H, Gu X. Correlation between drug 32. dissolution and polymer hydration: a study using texture analysis. Int J Pharm 2007;342:18-25
- 33. Jamzad S, Tutunji L, Fassihi R. Analysis of macromolecular changes and drug release from hydrophilic matrix systems. Int J Pharm 2005;292:75-85
- 34. Yang LB, Johnson B, Fassihi R. Determination of continuous changes in the gel layer thickness of poly(ethylene oxide) and HPMC tablets undergoing hydration: a texture analysis study. Pharm Res 1998;15:1902-6
- 35. Pavli M, Kristl J, Dolenc A, et al. The reflection of the texture of swollen polymer matrix on the release of incorporated substance. E-Polymers 2009 no. 017
- 36. Fukushima E, Roeder SBW. Experimental pulse NMR. A nuts and bolts approach. Perseus Publishing, Cambridge, Massachusetts; 1981
- Abragam A. Principles of nuclear magnetism. Oxford University Press, Oxford; 1961
- 38. Callaghan PT. Principles of nuclear magnetic resonance microscopy. Oxford University Press, New York; 1991
- 39. Meiboom S, Gill D. Modified spin-echo method for measuring nuclear relaxation times. Rev Sci Instr 1958;29:688-91
- Stejskal EO, Tanner JE. Spin diffusion 40. measurements: spin echoes in the presence of a time-dependent field gradient. J Chem Phys 1965;42:288-92
- 41. Morris KF, Johnson CS Jr. Diffusion-ordered two-domensional nuclear magnetic resonance spectroscopy. J Am Chem Soc 1992;114:3139-41
- 42. Schmidt-Rohr K, Spiess HW. Multidimensional solid-state NMR in polymers. Academis Press Ltd, London;
- 43. Henkelman RM, Huang X, Xiang QS, et al. Quantitative interpretation of magnetization transfer. Magn Reson Med 1993;29:759-66
- 44. Grad J, Bryant RG. Nuclear magnetic cross-relaxation spectroscopy. J Magn Reson 1990;90:1-8
- 45. Samoilenko AA, Artemov DY, Sibeldina LA. Formation of sensitive layer in experiments on NMR subsurface imaging of solids. JETP Lett 1988;47:417-19

- Hennig J, Nauerth A, Friedburg H. RARE imaging: a fast method for clinical MR. Magn Reson Med 1986;3:823-33
- Norris DG, Bornert P, Reese T, et al. On the application of ultra-fast RARE experiments. Magn Reson Med 1992;27:142-64
- Sederman AJ, Mantle MD, Gladden LF. Single excitation multiple image RARE (SEMI-RARE): ultra-fast imaging of static and flowing systems. J Magn Reson 2003:161:15-24
- Chen YY, Hughes LP, Gladden LF, et al. Quantitative ultra-fast MRI of HPMC swelling and dissolution. J Pharm Sci 2010;99:3462-72
- Gravina S, Cory DG. Sensitivity and resolution of constant-time imaging. J Magn Reson B 1994;104:53-61
- Beyea SD, Balcom BJ, Prado PJ, et al. Relaxation time mapping of short T2\* nuclei with single-point imaging (SPI) methods. J Magn Reson 1998;135:156-64
- Hills PB. The proton exchange cross-relaxation model of water relaxation in biopolymer systems. Mol Phys 1992;76:489-508
- Gao P, Fagerness PE. Diffusion in HPMC gels. I. Determination of drug and water diffusivity by pulsed-field-gradient spin-echo NMR. Pharm Res 1995;12:955-64
- Masaro L, Ousalem M, Baille WE, et al. Self-diffusion studies of water and poly (ethylene glycol) in solutions and gels of selected hydrophilic polymers. Macromolecules 1999;32:4375-82
- 55. Ferrero C, Massuelle D, Jeannerat D, et al. Towards elucidation of the drug release mechanism from compressed hydrophilic matrices made of cellulose ethers. I. Pulse-field-gradient spin-echo NMR study of sodium salicylate diffusivity in swollen hydrogels with respect to polymer matrix physical structure. J Control Release 2008:128:71-9
- Katzhendler I, Mader K, Friedman M. Structure and hydration properties of hydroxypropyl methylcellulose matrices containing naproxen and naproxen sodium. Int J Pharm 2000;200:161-79
- Katzhendler I, Mader K, Azoury R, et al. Investigating the structure and properties of hydrated hydroxypropyl methylcellulose and egg albumin matrices

- containing carbamazepine: EPR and NMR study. Pharm Res 2000;17:1299-308
- 58. Fyfe CA, Blazek-Welsh AI. Quantitative NMR imaging study of the mechanism of drug release from swelling hydroxypropylmethylcellulose tablets. J Control Release 2000;68:313-33
- Direct measurements of drug position during the swelling process followed by 19F MRI.
- 59. Brandl F, Kastner F, Gschwind RM, et al. Hydrogel-based drug delivery systems: comparison of drug diffusivity and release kinetics. J Control Release 2010:142:221-8
- Yasuda H, Lamaze CE. Permselectivity of solutes in homogeneous water-swollen polymer membranes. J Macromol Sci Phys 1971;B5:111-34
- 61. Korsmeyer RW, von Meerwall E, Peppas NA. Solute and penetrant diffusion in swellable polymers. II. Verification of theoretical models. J Polym Sci 1986;24:409-34
- Ruan RR, Long Z, Song A, et al. 62. Determination of the glass transition temperature of food polymers using low field NMR. Lebensm Wiss Technol 1998;31:516-21
- 63. Ruan R, Long Z, Chen P, et al. Pulse NMR study of glass transition in maltodextrin. J Food Sci 1999;64:6-9
- Hills BP, Pardoe K. Proton and 64. deuterium NMR studies of the glass transition in a 10% water-maltose solution. J Mol Liq 1995;63:229-37
- Calucci L, Forte C, Ranucci E. Water/polymer interactions in poly (amidoamine) hydrogels by 1H nuclear magnetic resonance relaxation and magnetization transfer. J Chem Phys 2008;129:064511
- Baumgartner S, Lahajnar G, Sepe A, et al. Investigation of the state and dynamics of water in hydrogels of cellulose ethers by 1H NMR spectroscopy. AAPS PharmSciTech 2003;3: no. 36
- Baumgartner S, Lahajnar G, Sepe A, et al. Quantitative evaluation of polymer concentration profile during swelling of hydrophilic matrix tablets using 1H NMR and MRI methods. Eur J Pharm Biopharm 2005;59:299-306
- Mikac U, Sepe A, Kristl J, et al. A new approach combining different MRI



- methods to provide detailed view on swelling dynamics of xanthan tablets influencing drug release at different pH and ionic strength. J Control Release 2010;145:247-56
- A new MRI approach for accurate determination of the penetration, swelling and erosion fronts.
- Fyfe CA, Blazek AI. Investigation of hydrogel formation from hydroxypropylmethylcellulose (HPMC) by NMR spectroscopy and NMR imaging techniques. Macromolecules 1997:30:6230-7
- Laity PR, Mantle MD, Gladden LF, et al. Magnetic resonance imaging and X-ray microtomography studies of a gel-forming tablet formulation. Eur J Pharm Biopharm 2010;74:109-19
- Vittadini E, Dickinson LC, Chinachoti P. NMR water mobility in xanthan and locust bean gum mixtures: possible explanation of microbial response. Carbohydr Poly 2002;49:261-9
- Dahlberg C, Fureby A, Schuleit M, et al. Polymer mobilization and drug release during tablet swelling. A 1H NMR and NMR microimaging study. J Control Release 2007;122:199-205
- Richardson JC, Bowtell RW, Mader K, et al. Pharmaceutical applications of magnetic resonance imaging (MRI). Adv Drug Deliv Rev 2005;57:1191-209
- An excellent review of pharmaceutical applications of MRI.
- Nott KP. Magnetic resonance imaging of tablet dissolution. Eur J Pharm Biopharm 2010;74:78-83
- This article provides a review of MRI measurements and instrumentations that are relevant to pharmaceutical applications.
- Mantle MD. Quantitative magnetic resonance micro-imaging methods for pharmaceutical research. Int J Pharm 27 Nov 2010. [Epub ahead of print]. Doi:10.1016/j.ijpharm.2010.11.035
- Narasimhan B, Snaar JEM, Bowtell RW, et al. Magnetic resonance imaging analysis of molecular mobility during dissolution of poly(vinyl alcohol) in water. Macromolecules 1999;32:704-10
- Snaar JEM, Bowtell R, Melia CD, et al. Self-diffusion and molecular mobility in PVA-based dissolution-controlled systems for drug delivery. Magn Reson Imaging 1998;16:691-4

- 78 Tajiri T, Morita S, Sakamoto R, et al. Release mechanisms of acetaminophen from polyethylene oxide/polyethylene glycol matrix tablets utilizing magnetic resonance imaging. Int J Pharm 2010:395:147-53
- 79 Tritt-Goc J, Pislewski N. Magnetic resonance imaging study of the swelling kinetics of hydroxypropylmethylcellulose (HPMC) in water. J Control Release 2002:80:79-86
- 80. Butler J, Nott K. Using low-field MRI to improve tablet dissolution testing. Tablets Capsules Jan 2010 Available at: http://www.tabletscapsules.com/ Content/getArticle.aspx? ItemID=7e3444d2-96be-4afd-ba 58-39a5d8255f90&Subject= Dissolution+testing
- Therien-Aubin H, Zhu XX, Ravenelle F, et al. Membrane formation and drug loading effects in high amylose starch tablets studied by NMR imaging. Biomacromolecules 2008;9:1248-54
- 82. Therien-Aubin H, Baille WE, Zhu XX, et al. Imaging of high-amylose starch tablets. 3. Initial diffusion and temperature effects. Biomacromolecules 2005;6:3367-72
- 83 Baille WE, Malveau C, Zhu XX, et al. NMR imaging of high-amylose starch tablets. 1. Swelling and water uptake. Biomacromolecules 2002;3:214-8
- Malveau C, Baille WE, Zhu XX, et al. NMR imaging of high-amylose starch tablets. 2. Effect of tablet size. Biomacromolecules 2002:3:1249-54
- Abrahmsen-Alami S, Koorner A, Nilsson I, et al. New release cell for NMR microimaging of tablets swelling and erosion of poly(ethylene oxide). Int J Pharm 2007;342:105-14
- 86. Fyfe CA, Grondey H, Blazek-Welsh AI, et al. NMR imaging investigations of drug delivery devices using a flow-through USP dissolution apparatus. J Control Release 2000;68:73-83
- 87. Bowtell R, Sharp JC, Peters A, et al. NMR microscopy of hydrating hydrophilic matrix pharmaceutical tablets. Magn Reson Imaging 1994:12:361-4
- Rajabi-Siahboomi AR, Bowtell RW, Mansfield P, et al. Structure and behavior in hydrophilic matrix sustained release dosage forms: 4. Studies of water

- mobility and diffusion coefficient in the gel layer of HPMC tablets using NMR imaging. Pharm Res 1996;13:376-80
- 89. Raiabi-Siahboomi AR, Bowtell RW, Mansfield P, et al. Structure and behaviour in hydrophilic matrix sustained release dosage forms: 2. NMR-imaging studies of dimensional changes in the gel layer and core of HPMC matrices undergoing hydration. J Control Release 1994:31:121-8
- 90. Fyfe CA, Blazek AI. Complications in investigations of the swelling of hydrogel matrices due to the presence of trapped gas. J Control Release 1998;52:221-5
- 91. Madhu B, Hjartstam J, Soussi B. Studies of the internal flow process in polymers by 1H NMR microscopy at 500 MHz. J Control Release 1998;56:95-104
- Kojima M, Ando S, Kataoka K, et al. Magnetic resonance imaging (MRI) study of swelling and water mobility in micronized low-substituted hydroxypropylcellulose matrix tablets. Chem Pharm Bull 1998;46:324-8
- Kojima M, Nakagami H. Investigation of water mobility and diffusivity in hydrating micronized low-substituted hydroxypropyl cellulose, hydroxypropylmethyl cellulose, and hydroxypropyl cellulose matrix tablets by magnetic resonance imaging (MRI). Chem Pharm Bull 2002;50:1621-4
- Tajarobi F, Abrahmsen-Alami S, Carlsson AS, et al. Simultaneous probing of swelling, erosion and dissolution by NMR-microimaging - Effect of solubility of additives on HPMC matrix tablets. Eur J Pharm Sci 2009;37:89-97
- Hyde TM, Gladden LF. Simultaneous measurement of water and polymer concentration profiles during swelling of poly(ethylene oxide) using magnetic resonance imaging. Polymer 1998;39:811-19
- 96. Goerke U, Chamberlain AHL, Crilly EA, et al. Model for water transport into powdered xanthan combining gel swelling and vapor diffusion. Phys Rev E 2000;62:5353-9
- Marshall P, Snaar JEM, Ng YL, et al. A novel application of NMR microscopy: measurement of water diffusion inside bioadhesive bonds. Magn Reson Imaging 2001:19:487-8
- Metz H, Mader K. Benchtop-NMR and 98. MRI - a new analytical tool in drug



#### Using quantitative magnetic resonance methods to understand better the gel-layer formation on polymer-matrix tablets

- delivery research. Int J Pharm 2008:364:170-5
- This paper presents the commercial low-cost bench-top MRI system and its applicability for pharmaceutical applications.
- 99 Strubing S, Abboud T, Contri RV, et al. New insights on poly(vinyl acetate)-based coated floating tablets: characterisation of hydration and CO2 generation by benchtop MRI and its relation to drug release and floating strength. Eur J Pharm Biopharm 2008;69:708-17
- 100. Strubing S, Metz H, Mader K. Characterization of poly(vinyl acetate) based floating matrix tablets. I Control Release 2008:126:149-55
- 101. Malaterre V, Metz H, Ogorka J, et al. Benchtop-magnetic resonance imaging (BT-MRI) characterization of push-pull osmotic controlled release systems. J Control Release 2009;133:31-6
- 102. Nott KP. Visualising tablet dissolution. Masurement of hydration and drug release. GIT Lab J 2008;9-10:42-3
- Kaunisto E, Abrahmsen-Alami S, Borquist P, et al. A mechanistic modelling approach to polymer

- dissolution using magnetic resonance microimaging. J Control Release 2010;147:232-41
- 104. Karakosta E, Jenneson PM, Sear RP, et al. Observations of coarsening of air voids in a polymer-highly-soluble crystalline matrix during dissolution. Phys Rev E 2006;74:011504
- 105. Ercken M, Adriaensens P, Reggers G, et al. Use of magnetic resonance imaging to study transport of methanol in poly (methyl methacrylate) at variable temperature. Macromolecules 1996;29:5671-7
- 106. Peppas NA, Wu JC, von Meerwall ED. Mathematical modeling and experimental characterization of polymer dissolution. Macromolecules 1994;27:5626-38
- 107. Devotta I, Premnath V, Badiger MV, et al. On the dynamics of mobilization in swelling-dissolving polymeric systems. Macromolecules 1994;27:532-9
- 108. Idiyatullin D, Corum C, Park JY, et al. Fast and quite MRI using a swept radiofrequency. J Magn Reson 2006;181:342-9

- 109. McConnell EL, Fadda HM, Basit AW. Gut instincts: explorations in intestinal physiology and drug delivery. Int J Pharm 2008;364:213-26
- 110. Steingoetter A, Weishaupt D, Kunz P, et al. Magnetic resonance imaging for the in vivo evaluation of gastric-retentive tablets. Pharm Res 2003;20:2001-7

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